A deeper look at the relationship between root C pools and the vertical distribution of soil C pools under maize and reconstructed prairie

1 Introduction

Prairie-formed mollisols support some of the world’s most productive agriculture, but declines in levels of soil organic matter threaten the perpetuity of this production. Soil organic matter losses coincide with a shift from perennial plant systems to annual cropping systems which introduced frequent tillage, subsurface drainage, and differences in organic matter inputs, including considerably different rooting systems. The effects of changes in aboveground processes such as increased soil disturbance and aeration, addition of fertilizers, and changes in residue amount and quality have been widely studied (cite, cite, cite). The role played by changes in rooting systems, on the other hand, is difficult to study and has received less attention.

In this paper, we distinguish between a root C pool (C found in any material that can still be visually identified as a root) and a soil organic C pool (the rest of the soil organic C). Root growth allows the placement of plant tissue directly into the soil, creating a root C pool as deep as the rooting system goes. Some studies suggest that root C pool size and soil organic C pool size have a direct relationship and most soil organic matter is derived from roots (Balesdent and Balabane 1996, Kong and Six 2010). This would mean that a change in root inputs, such as switching from annual to perennial systems, would have a direct impact on soil organic matter even deep into the soil profile. However, very few direct comparisons of annual and perennial rooting systems have been made and our understanding of soil C dynamics decreases as depth increases.

On average, half of the world’s soil C is found below 20 cm (Rumpel and Kogel-Knabner, 2011). However, only 30% of the world’s roots are found below 20 cm (Jobbagy and Jackson, 2000). In the Central US, this phenomenon was observed as early as 1935 when Weaver found 41-74% of the total soil organic matter, but only 23-29% of the total roots in a tallgrass prairie were found below 20 cm. Similarly, Gill and colleagues (1999) found 77% of total soil organic matter and only 43% of the total root mass below 15 cm in a shortgrass steppe. Although this disproportionate relationship between root and soil C distribution is well known, and has been for some time, no widely accepted explanation exists to justify the magnitude of difference between the amount of C in the root pool and the amount of C in the soil pool (Gill et al., 1999, Rumpel and Kogel-Knabner 2011).

Many factors interact to determine how much C is transferred between pools and how much C remains in a particular pool. Temperature, moisture, and O2 availability are the most important environmental variables controlling the rate of decomposition (Gill and Burke, 2002) and soil texture and existing soil C levels determine the length of time C remains in the soil (Six et al. ????). The C:N ratio of the organic matter being decomposed also plays a key role in both the rate of decomposition and the fate of the decomposed organic matter, with higher C:N ratios leading to slower decomposition (Silver and Miya 2001) and fewer microbial by-products (Cortrufo 2015). Temperature, moisture, O2, soil texture, and soil C levels all vary with depth and contribute to partial explanations of the size discrepancy between root and soil C pools. However, previous studies which measured roots and/or organic matter with depth have neglected to report the change of root C:N ratio with depth (Tufekcioglu et al. 2003, Beniston et al. 2014). Carbon:N ratio differences between maize and prairie root C pools are also unknown. A more-detailed look at properties of root C pools is greatly needed.

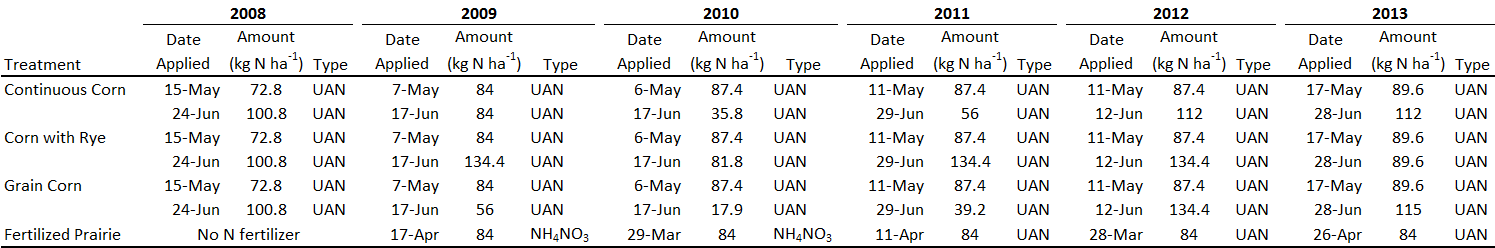
We examined the belowground reconstruction of native vegetation on a Mollisol after >100 years of annual cropping to gain a unique perspective on characteristics of root inputs that would not necessarily be noticed in established prairie systems, but that contribute to dynamics of the belowground ecosystem. We examined differences between maize and reconstructed prairie root pools down to one meter to serve two separate, but related, purposes: 1) Inform our understanding of the impact and lasting effect of shifting millions of acres from perennial to annual vegetation and; 2) Contribute to an explanation of why levels of soil organic C found below 20 cm are greater than expected. In comparing the root C pool of a reconstructed prairie system to the root C pool of a maize cropping system we asked the following questions: 1) How does the quantity, distribution, and C:N ratio of the root C pool differ with depth and between these native perennial and non-native annual ecosystems? 2) What do these differences in inputs tell us about the historical belowground ecosystem under which these soils developed and the systems under which these soils continue to change?

2 Materials and Methods

*2.1 Site Conditions and Experimental Design*

We conducted the experiment in Boone County, IA, USA on the Iowa State University Agronomy and Agricultural Engineering Research Farm (41°55′N, 93°45′W). Soils at the site were primarily Webster silty clay loam (ﬁne-loamy, mixed, superactive, mesic Typic Endoaquoll) and Nicollet loam (ﬁne-loamy, mixed, superactive, mesic Aquic Hapludoll). The 60-year mean growing season precipitation 11 km from the site was 720 mm. Prior to initiation of the ﬁeld experiment in 2008, the site was used for maize and soybean production and was planted with soybean in 2007. Soil sampling to 15 cm in November 2007 indicated mean soil pH was 6.7, mean soil C concentration (via dry combustion analysis) was 30 g kg-1, mean extractable phosphorus concentration (via Bray-1 procedure) was 11 mg kg-1, and mean extractable potassium (via Mehlich-3 procedure) was 141 mg kg-1.

Experimental plots were 27 m x 61 m and were arranged as a spatially balanced complete block design (van Es *et al*., 2007). The three cropping systems used in this study were continuous maize with annual grain and stover removal (hereafter maize), reconstructed multispecies prairie with annual aboveground biomass removal (hereafter unfertilized prairie), and N-fertilized reconstructed multispecies prairie with annual aboveground biomass removal (hereafter fertilized prairie). All of the treatments were managed without tillage. Conventional farm machinery was used for planting, fertilization, crop protection, and harvest operations. Herbicides were not used in the prairie systems except for a small number of spot treatments for Canada thistle (*Circium canadense*) control, and the timing and frequency of herbicide use in the annual cropping systems varied among treatments. Nutrient management also varied among all treatments (Table 1).

Both prairie treatments were sown on 19 May 2008 with the same custom seed mix obtained from Prairie Moon Nursery (Winona, MN, USA) that contained 31 species, including C3 and C4 grasses and leguminous and non-leguminous forbs. All species were perennial and sourced from within 240 km of the experiment site. The composition of the seed mix by weight was 12% C3 grasses, 56% C4 grasses, 8% legumes, and 24% non-leguminous forbs. A detailed description of the prairie plant community compositions can be found in Jarchow and Liebman (2013). The Table 1. N fertilizer amount, type, and date applied for all COBS treatments. Need to update.

fertilized prairie treatment received no fertilizer in 2008 (the establishment year), and was fertilized at a rate of 84 kg N ha-1 year-1 in all subsequent years. This fertilizer rate was chosen because it was similar to the maximum rate of pre-planting N fertilization recommendedfor maize (Blackmer *et al*., 1997) and the expected N removal in the harvested biomass of perennial grasses grown in the area (Heggenstaller *et al*., 2009). Fertilization timing can be found in Table 1.

The maize used was a 104-day relative maturity hybrid with transgenes for glyphosate resistance, corn borer (*Ostrinia nubilalis*) resistance, and corn rootworm**(***Diabrotica* spp.) protection (Agrigold 6325 VT3). Maize was planted following standard practices (Abendroth *et al*., 2011) in rows spaced 76 cm apart at 79,500 seeds ha-1 on 6 May 2010 and 82,500 seeds ha-1 on 11 May 2011. Fertilizer rates and types can be found in Table 1. Rates of N added after planting were based on results of late-spring tests of soil nitrate-N concentration (Blackmer *et al*., 1997). All N was applied as urea-ammonium nitrate (32% N). An unfertilized maize treatment was not included in the experiment because the effects of N fertilizer on maize have been extensively studied and modeled, with N fertilization leading to greater biomass and higher grain yields (Cerrato and Blackmer 1990; Sawyer *et al*., 2006).

*2.2 Data Collection*

*2.2a Soil Collection*

Soil cores were taken to 1 m depth in all plots each year using a hydraulic soil probe (Giddings Machine Co., Windsor, CO, USA) after all crops were harvested. Sampling occurred by replicate block from 31 October-25 November 2008, 9-11 November 2009, 25-28 October 2010, 28-31 October 2011, 16-17 October 2012, and 7-11 October 2013.

In 2008, two cores were taken per plot. A 0-30 cm fraction was taken with a 10.2 cm internal diameter soil probe; the 30-100 cm fractions of the cores was taken within the same hole as the 0-30 cm fraction, but with a smaller soil probe. In Blocks 1 and 4, the internal diameter of the core was 6.0 cm. In Blocks 2 and 3, the internal diameter of the core was 5.2 cm.

In 2009 and 2010, four cores were taken per plot. The 0-30 cm fraction of the cores was taken with a 10.2 cm internal diameter soil probe; the 30-100 cm fraction of the cores were taken directly below the 0-30 cm fraction with a 5.1 cm internal diameter probe. In 2011-2013, four cores were taken per plot, and the entire core was taken with a 5.1 cm internal diameter probe.

Soil cores were ultimately divided into three or five depth increments. In 2008, depth increments were 0-30 cm, 30-60 cm, and 60-100 cm. In 2009-2013 depth increments were 0-5 cm, 5-15 cm, 15-30, cm, 30-60 cm, and 60-100 cm. Following division and extraction from the field, soil cores were stored at 5°C until processing was initiated.

Each year, 60-100 g of root-free soil was removed from each depth increment, air-dried, and archived in airtight containers at room temperature. In 2008 and 2013, this soil was ground on a roller-mill and organic C content was determined by catalytic oxidation and CO2 measurement with NDIR in an Elementar TOC Cube at Brookside Laboratories, Inc. (New Bremen, Ohio).

*2.2b Root Pool Collection*

It is important to note that two separate sets of root pools samples were collected. The first, described in this section, was used to track changes in the root C pools over all six years and the second, described in section 2.2c, was used to quantify annual root C contributions in 2010 and 2011.

Root extraction from the soil began by washing the soil samples described in 2.2a in wire mesh tubes (0.28 mm mesh) for 3 h in an elutriator (Wiles et al., 1996). Roots were removed from the remaining soil by suspending the air-dried sample in water and collecting the roots, which floated, with sieves followed by manually removing any remaining non-root material that was present in the samples. Any plant crowns that were present in the samples were removed and were not considered to be root biomass. Roots were then dried at 70°C for at least 4 h before being weighed. All root samples were ground to 2 mm with a centrifugal mill and concentrations of C and N were determined by combustion analysis at the Soil and Plant Analysis Laboratory at Iowa State University (Ames, IA, USA).

*2.2c In-season Root Growth*

In 2010 and 2011, root biomass was measured with an in-situ growth core approach (Neill 1992) to capture only those roots growing within the measurement year. After fall harvest in 2009 and 2010, eight 10.2-cm-diam soil cores were taken to 30 cm depth in each plot and brought to the laboratory. Holes created in the field were held open during the winter by capped 10.2 cm PVC piping. In the laboratory, cores were divided into 10 cm sections and virtually all roots were removed by hand. Soil was stored in intact cores at 30°C for the first year of the experiment and 4 °C in sealed plastic bags for second year of the experiment. The differences in storage conditions did not have an apparent effect on the outcome of the experiment. At the end of winter while plants were still dormant, the root-free soil was returned to its original location in the field in 10 cm depth increments. Soil was packed to imitate the surrounding bulk density, approximately 1.4 g cm-3. Root-free zones were located randomly within prairie plots and at 20 cm from maize rows. Eight root-free areas were situated within each plot, allowing duplicate sampling at four time points throughout the growing season. Two 4-cm-diam soil cores were taken within each 10.2-cm-diam root-free area to a 30 cm depth at each root sampling date. Bulk soil was washed from the roots with water using a soil elutriator (Wiles *et al*., 1996), roots were dried at 60° C for 24 hours, non-root biomass was removed from the roots by hand, and roots were weighed.

*2.3 Data Analysis*

Root pool mass for the entire meter depth was calculated by summing together the root mass for each depth increment of an entire core and whole core root masses between treatments were compared within each year using contrasts within a linear mixed effect model in the R package *nlme* (Pinheiro et al. 2013). Treatment differences within depths within years and differences between treatments within depths within years for root biomass were also made with contrasts with linear mixed effects models, but proc glimmix in SAS (SAS Institute, 2011) was used.

Because root mass in 2008 was measured at three increments (0-30 cm, 30-60 cm, and 60-100 cm) instead of five increments (0-5 cm, 5-15 cm, 15-30 cm, 30-60 cm, and 60-100 cm), 2008 root mass for 2008 0-5 cm, 5-15 cm, and 15-30 cm was estimated by multiplying the average 2009-2013 proportion by the 2008 0-30 cm increment. No important comparisons were made using this estimated data, but the data were used as a starting point for graphing C:N ratios in different depth increments and fitting curves to root accumulation. Carbon:N ratios were compared between treatments within years within depths and between years within treatments within depths using proc glimmix in SAS.

Root mass measured at the end of the each growing season was subset by depth increment and each subset was fit by both a logistic model and a linear model for each plot. Logistic models and linear models were compared against each other using Akaike’s Criterion (AIC) and the model with the lowest AIC was chosen. The AIC was not vastly different for any of the comparisons, but the logistic model had the best fit for every depth (Appendix). Model fits and comparisons were done using the R package *nlme* (Pinheiro et al. 2013).

The first derivative of the logistic model was used to calculate the daily rate of accumulation. Parameters from the logistic model were used to predict both amount and rate of accumulation for each day for each depth in each plot of the experiment. These predictions were averaged for each treatment and plotted. The annual mean rate was calculated by averaging accumulation rates across each growing season for each depth in each plot. Comparisons of rates between treatments within depths and within years and comparisons of rates between depths within treatments within years were made with proc glmmix in SAS.

In-situ root measurements in 2010 and 2011 combined with differences in root pool masses at 30 cm over these years were used to calculate a root turnover constant (k) and root mean residence time (mrt) using the equations k = loss/pool and mrt = 1/k . Root pool loss during each year was calculated as the difference between the mass accumulated during that year and the gain measured by in-situ growth cores. The root mass measured at the end of each year was the pool value.

Because root samples were not taken at equivalent depth increments, splines were fit to the existing data and integrated by 5 cm depths to create accurate visualization of root and soil organic C distribution in the soil profile.

3 Results

Table 1. Soil characteristics. soil C, soil N, clay, sand, silt, pH (to be created)

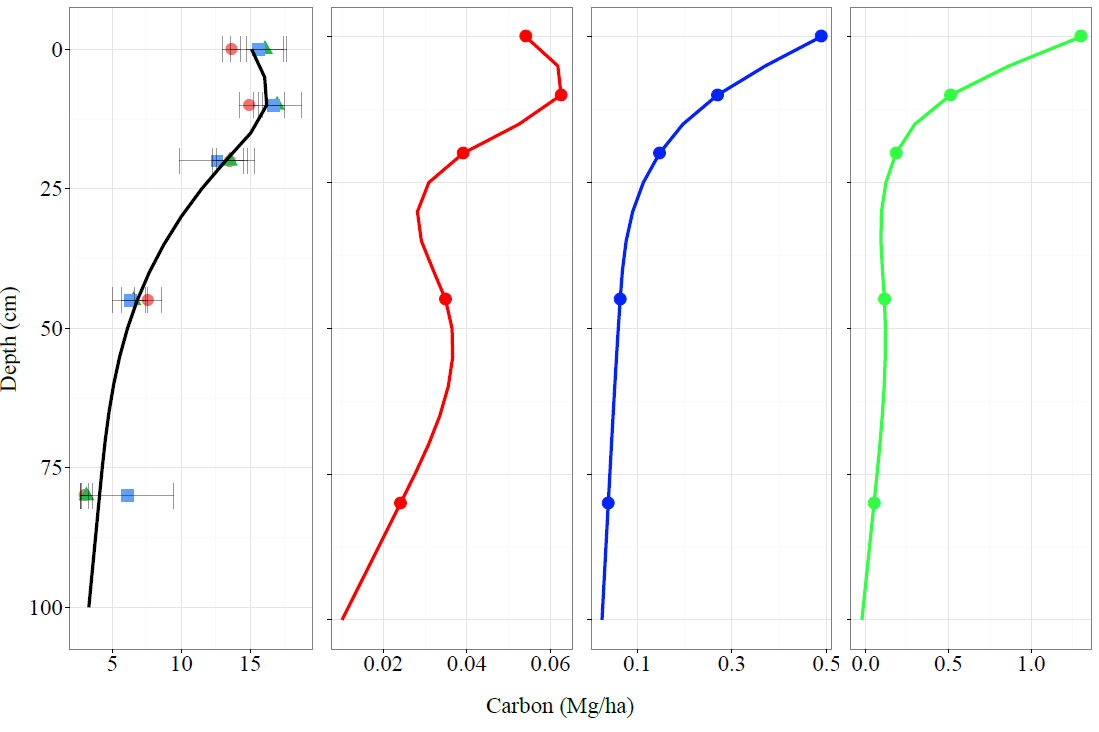


Figure 1. From left to right: total soil carbon, maize root carbon, fertilized prairie root carbon, unfertilized prairie root carbon at the end of the study (2013). Different x-axes scales are used to emphasize similarities and differences in distribution patterns, not absolute mass amounts (see Fig 2).

The amount of total organic C found in the soil 6 years after establishment of the experiment was not different among treatments at any depth (Fig 1), nor was it different from initial total organic C levels measured at the beginning of the experiment (data not shown). Half of the total soil organic C was found to be below 20 cm (Table 2). The pattern of vertical soil C distribution reflected the pattern of maize root distribution, not prairie root distribution (Fig 1).

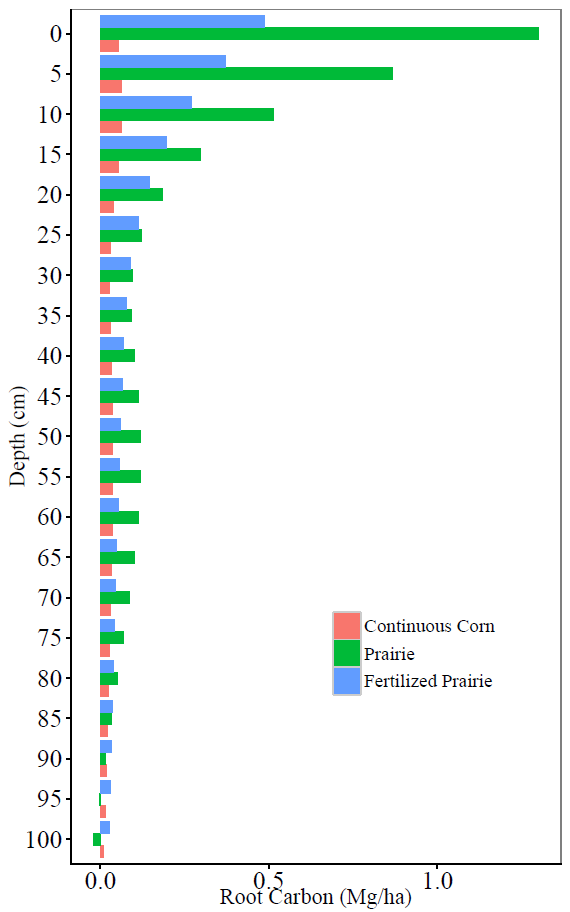


Figure 2. Absolute difference in root C pools six years after prairie establishment.

Six years after the establishment of the experiment, the unfertilized prairie root C pool was almost 6 times greater than the maize root C pool and the fertilized prairie root C pool was 3.5 times greater than the maize root C pool over a 1 m depth. Twenty-eight percent of the unfertilized prairie root C pool, 37% of the fertilized prairie root C pool and 62% of the maize root C pool was found below 20 cm (Fig 2, Table 2).

Table 2. Root pool and soil organic C found above and below 20 cm.



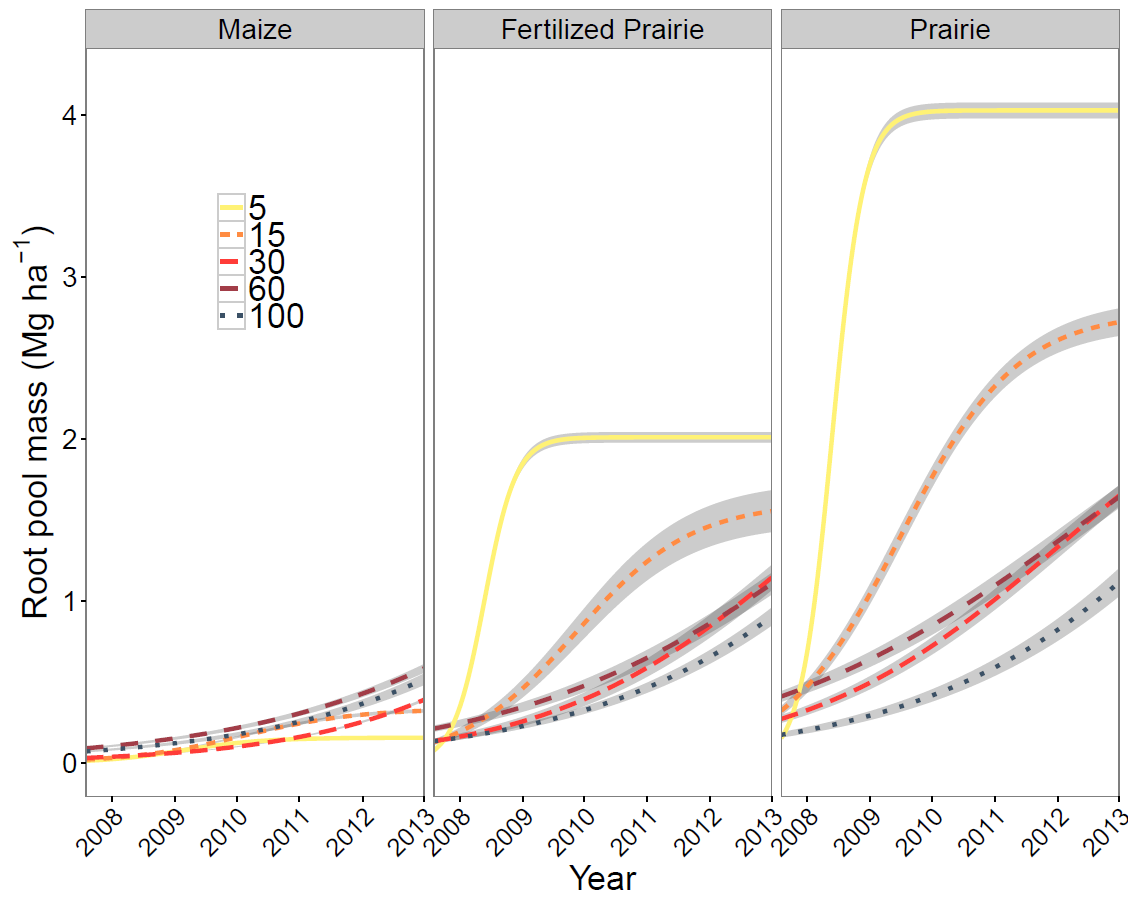


Figure 3. Accumulation of root pool mass over six years at 0-5 cm, 5-15 cm, 15-30 cm, 30-60 cm, and 60-100 cm.

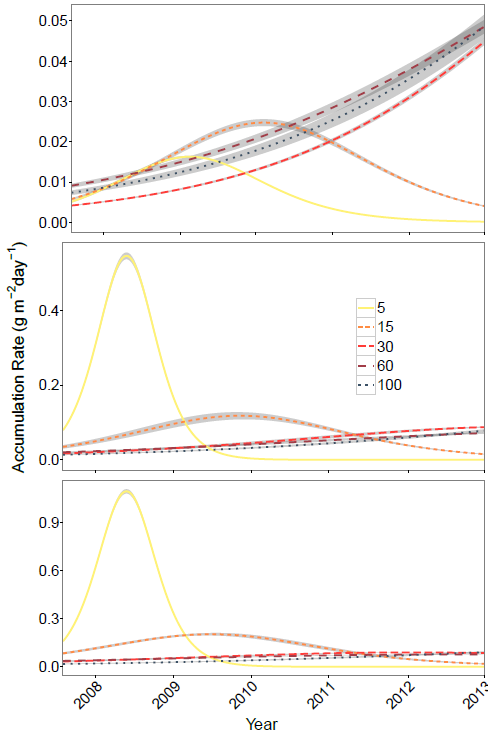


Figure 4. Rates of root pool mass accumulation over 6 years in continuous corn (top), fertilized prairie (middle) and unfertilized prairie (bottom) at 0-5 cm, 5-15 cm, 15-30 cm, 30-60 cm, and 60-100 cm. Please note different y-axes are used to emphasize similarities and differences in timing as well as make within treatment relationships easier to see.

Table 3. Root pool accumulation rates averaged across each growing season. Differences in lowercase letters indicate significant differences between depths within treatments within years (read up and down). Differences in uppercase letters indicate differences between treatments within depths within years (read left to right).



Prairie rooting systems were established sequentially in the soil profile from the top down. The top five cm of the root pool peaked in the first full year of growth and then reached an equilibrium during the second full year of growth with large year-to-year variability given the sensitivity of this thin surface layer to environmental conditions (Fig S1). The next soil layer, from 5-15 cm, experienced the greatest increase in root pool mass during the second full year of prairie growth, while the 15-30 cm and 30-60 cm depths didn’t reach peak rates of root pool accumulation until five and six years after establishment, with no indication of when accumulation will cease. In the unfertilized prairie, rates of root pool accumulation in the 60-100 cm of the soil in the sixth year were greater than all other depths with no sign of slowing down. Fertilized prairie also had a high rate of root pool accumulation at 60-100 cm in the sixth year with no sign of decreasing.

Maize root pool accumulation was almost always slower than prairie root pool accumulation with the exception of the top 5 cm after 2010, 60-100 cm before 2011 (not different from fertilized prairie), and a greater value in maize than unfertilized prairie at 30-60 cm in 2013. There was no difference in maize root pool accumulation among depths until 2011 when accumulation below 15 cm then began to exceed accumulation above 15 cm.

Table 3. Root turnover at 0-30 cm.

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trt input gain loss pool k mrt

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Prairie2010 367 104 263 748 0.35 2.8

Prairie2011 387 78 309 758 0.41 2.5

FertilizedPrairie2010 146 62 84 231 0.37 2.7

FertilizedPrairie2011 168 55 113 342 0.33 3.0

Maize2010 56 18 38 44 0.86 1.2

Maize2011 48 16 31 47 0.67 1.5

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Prairie roots had a mean residence time (mrt) of 2.75 years in the top 30 cm of the profile when averaged across treatments and years (2010 and 2011). Maize roots turned over twice as fast as prairie roots when averaged across treatments and years (Table 3).

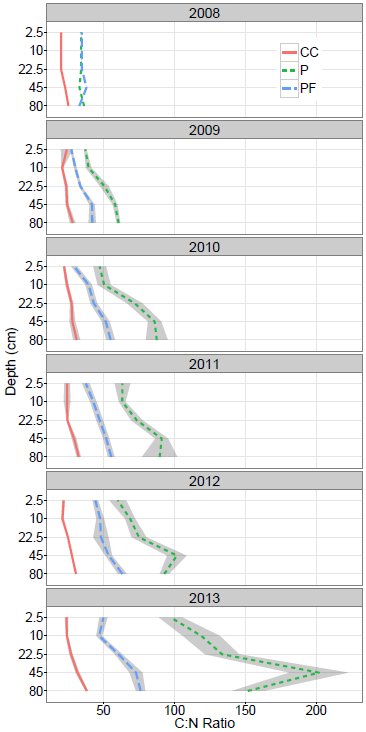


Figure 4. Root C:N ratios with depth over time.

Carbon:N ratios increased with depth in all treatments. Carbon to nitrogen ratios increased in both prairie treatments in every depth over time, although the increase in fertilized prairie was not always different between consecutive years. In all treatments, changes in C:N ratios were the result of both an increase in C content and a decrease in N content (data not shown). The maize root pool did not have an increase in C:N ratio over time.

4 Discussion

*4.1 Reconstruction of a prairie root C pool and implications for C contribution*

An increase in root pool C:N ratio with depth has not been previously reported in the literature and not previously considered when trying to determine why an unexplainably large amount of soil C is found at depth when compared to root distribution. It has been recently theorized that plant tissue becomes organic matter through two different pathways, 1) a dissolved organic carbon-microbial pathway whereby plant litter is first processed by soil microbes and eventually transported and stabilized in the soil matrix as microbial by-product, if the soil has the capacity to stabilize these compounds and; 2) a physical-transfer pathway whereby plant tissue is processed by soil microbes to its fullest extent, then remains in the soil functionally inert (Cortrufo 2015). In this study, the former pathway is more applicable to tissue dominated by non-structural compounds, such as that with lower C:N ratios found here at shallower depths, while the latter applies to tissue dominated by structural compounds, indicated by high C:N ratios in root tissue at deeper depth. Under this framework, root decomposition in our study would have resulted in a gradient of microbially-derived to physically-derived organic matter from the top down. In turn, this would mean that soil organic matter at the soil surface would be vulnerable to transport to deeper depth as dissolved organic C, while physically-transferred soil organic matter at depth would be relatively immobile. This is a possible mechanism by which the amount of soil organic C found at depth is disproportionately large compared to the size of the root C pool. These findings are consistent with evidence that the contribution of microbial- and not root-derived C increases with depth (Liang and Balser 2008, Rumpel and Kogel-Knabner, 2011).

Because the root pool is made up of a combination of new, mature, ageing, and dead roots, its increase in mass comes from root growth, live root retention, and inhibited root decomposition. The relatively quick rate of accumulation in the top 30 cm of soil was most likely a function of new root addition, which slowed as the system became more established. Slower increases at deeper depths than shallower depths may be indicative that accumulation there is more dependent upon the carryover of roots from previous years than at shallower depths.

By the sixth year of reconstructed prairie establishment, root C pool equilibrium was reached and prairies began making substantial annual contributions to the soil organic matter pool above 30 cm, although the fraction of organic matter that remained in the soil is unknown. This was indicated by the finding that the majority of prairie roots (75%) was found in this depth fraction, where mean residence time was measured to be 2.5-3 years. The prairie root C pool at 0-5 cm reached an equilibrium and began steady root turnover in the third year after establishment, as indicated by very low rates of accumulation, and was likely able to contribute material to the soil organic matter pool at this time. Other prairie restorations have also found soil organic matter accumulation to be most rapid closer to the soil surface (O’Brien et al. 2010, Omonode and Vyn 2006).

Annual prairie root inputs were not measured below 30 cm, so turnover rates could not be calculated. However, continuous increases in the root pool at depth due to root growth and retention indicates that root tissue loss to the soil was very low during this time and the mean residence time of roots at depth may greatly exceed those closer to the surface. This means that at depth, not only is the root C pool substantially smaller than near the surface, but root material also becomes available to the soil much more slowly than near the surface. Indeed, DuPont et al. (2014) found prairie roots five years after conversion to annual wheat.

The increase in C:N ratio with depth may have been due to difference in root age, even in maize. The deepest roots were the oldest roots (York, personal communication). The effect of time on increase of root C:N ratio was most obvious in prairie, which may have been a function of both maturing roots and the inclusion of dead roots in the root C pool measurements.

Nitrogen fertilization of prairies led to a smaller root pool at every depth, with lower rates of accumulation, and lower C:N ratios. However, fertilization did not affect the time until root systems were fully established or the turnover rate of roots in the top 30 cm. Differences between fertilized and unfertilized prairie show that the pattern of distribution was a function of nutrient availability and not a response to soil space conditions because fertilized prairie used half as much space as unfertilized prairie and still showed decreased accumulation above 30 cm over time.

*4.2 Quantity, distribution, and quality of root biomass differs in native perennial and non-native annual ecosystems*

It is possible that maize roots contribute more C to the soil than do prairie roots below a certain depth. Maize root C pools were much smaller than prairie root C pools, but faster turnover times and lower C:N ratios resulted in a greater proportion of the maize root C pool available for stabilization in the soil compared to the prairie root C pool. In the top 0-30 cm, the difference in mass between even the fertilized prairie and maize was too great to be overcome by faster turnover and greater carbon use efficiency, but the difference in root mass between the annual and perennial systems decreased with depth while the difference in C:N ratio increased and turnover times may have maintained the same relative relationship.

*4.3 What do these differences in inputs tell us about the historical belowground ecosystem under which these soils developed in comparison to the systems under which these soils continue to change?*

The experimental location was a site of cultivation under annual crops for over 100 years. We do not have a measurements of the pre-cultivation soil C profile, but other data from sites around the area (Guzman 2009, McGranahan et al. 2014) show that the soil C profile shifted from a pattern of having an exponential decrease in C with distance from the surface to a pattern of more uniform distribution of C with the highest point of C 10 cm below the surface. The loss of C in the soil surface after cultivation is well known and attributed to mass loss through soil erosion, increased mineralization of organic matter through tillage, and decreased belowground organic matter inputs. Less is documented about the change in soil carbon below 30 cm, but using a very robust dataset, Veenstra et al. (2015) found soil organic C to increase below 35 cm after 50 years in maize and soybean cropping systems in Iowa. Initial soil organic C measurements were made ~50 years after these soils had already been converted to annual systems, preventing comparison to soil organic C levels at depth under native vegetation, but these results still show that Mollisols can and do gain soil organic C at deeper depths under maize and soybean systems.

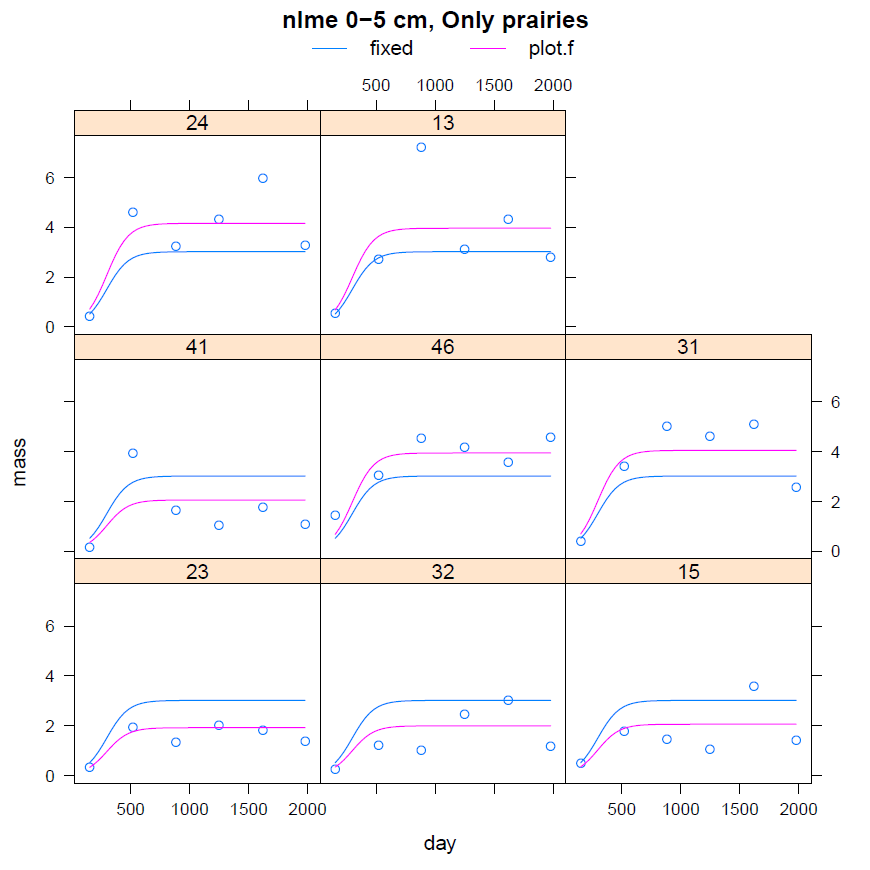
Our relatively short-term study of 6 years was not able to capture significant changes in soil C at any depth, but differences in quantity, distribution, and C:N ratios between the annual and perennial rooting systems we studied have important implications for how deep soil organic C may have changed and continues to change with the implementation of annual cropping systems. A large, structural-tissue-dominated root C pool with slow turnover, concentrated at shallow depths was replaced by a small, non-structural-tissue root pool with fast turnover evenly distributed in the soil profile. The difference in size between these two pools has long been obvious, but often misleading for comparisons related to C accounting because differences in root turnover and tissue C:N ratio are not taken into consideration. The data presented here in the context of recent organic matter formation theory suggest that while differences in root C pool and soil organic C relationships in maize and prairie above 20 cm are predominately controlled by root biomass amount, root biomass amount is less of a factor below 20 cm. While we have seen a dramatic decrease in soil C near the soil surface with conversion to annual crops, this is not necessarily true below 20 cm.

5 Conclusion

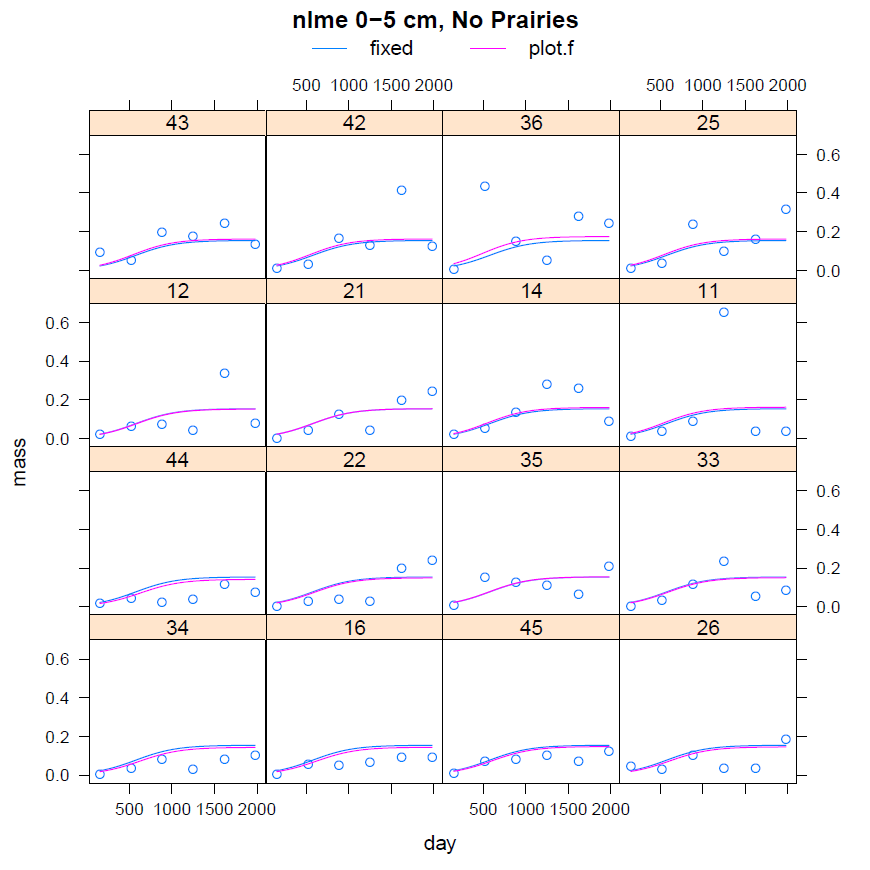
Soils are incredibly complex and biogeochemical processes which determine C storage happen over a long time and in environments that are difficult to study without causing disturbance. We have shown here that an increase in root C:N ratio with depth is a potentially important, and previously unconsidered, factor determining the distribution of C in the soil profile. This factor interacts with depth-determined differences in soil temperature, moisture, O2, texture, microbial communities, and existing soil C content and thus carries different significance in different environments. In our comparison of maize and reconstructed prairie systems, root pool C:N ratios may be an important enough factor to result in greater maize C contributions to soil organic matter than prairie C contributions to soil organic matter below 30 cm. In these and many other herbaceous systems, an increase in root C:N ratio with an increase in depth may in part explain why 50% of soil organic C is found below 20 cm while only 30% of root biomass is found in the same location. Elucidating the mechanisms behind soil C retention and addition is important as we strive to design systems which maintain and build soils that are productive and resilient. The role of roots and root composition, as well as the importance of soils below 20 cm should be carefully considered in such design.

6. Appendix

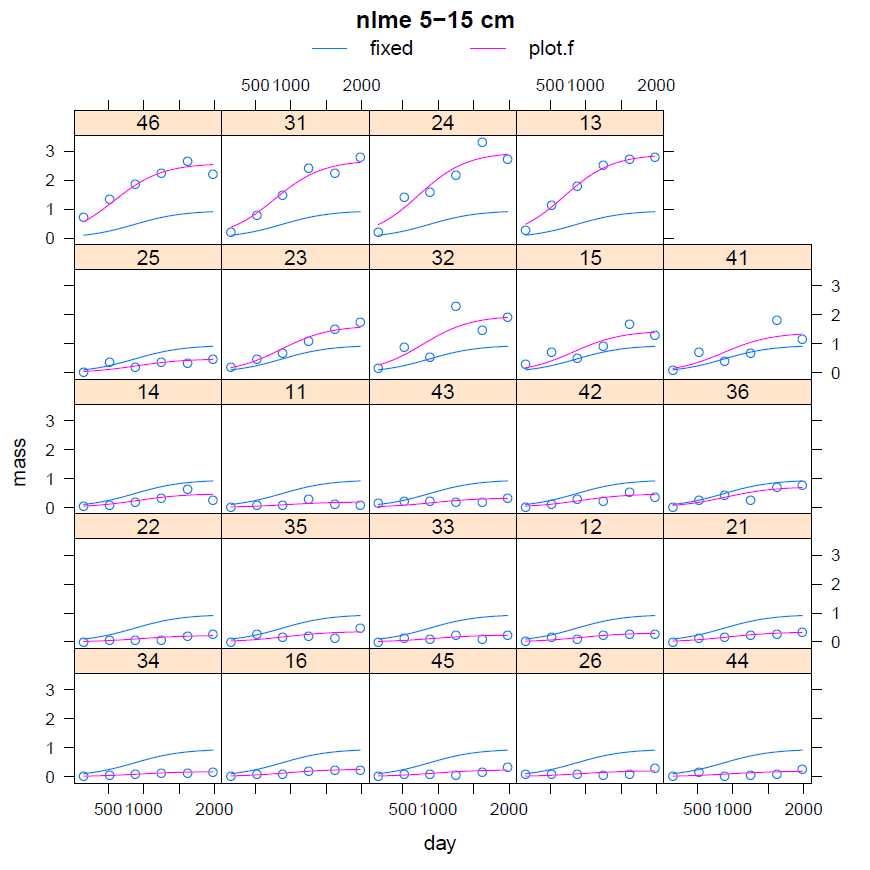
*Logistic curve fits used to generate predicted root accumulation for each depth. Each panel represents one experimental plot (number is plot number).*



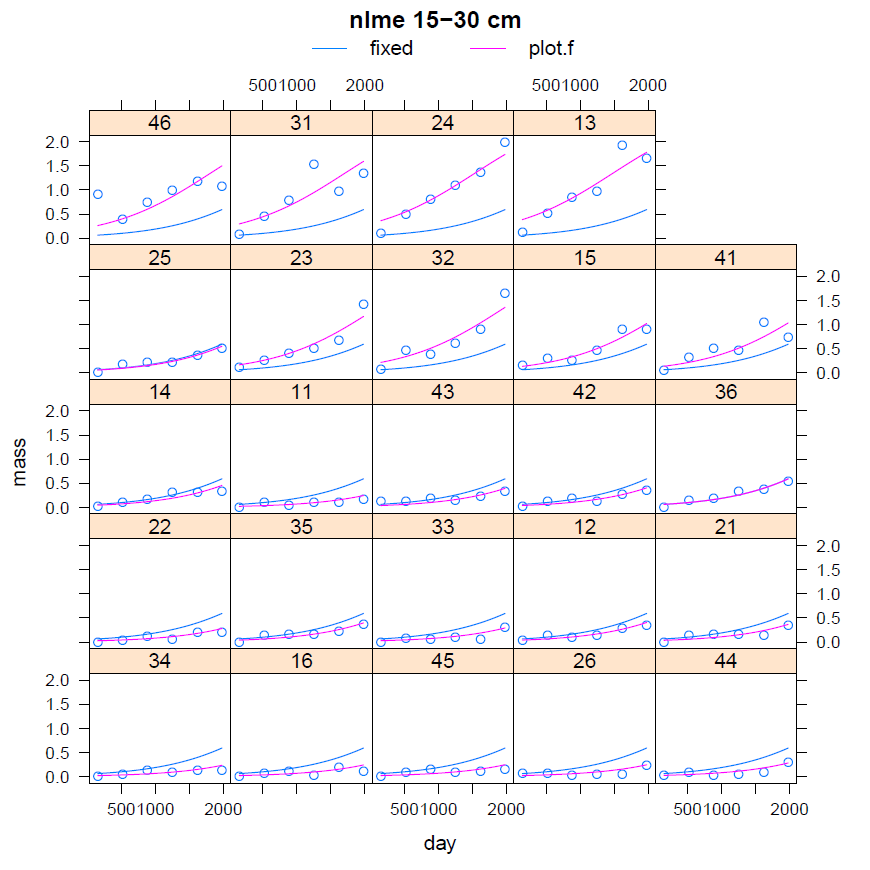
Appendix Fig 1. Fits of logistic curves to 0-5 cm prairie and unfertilized prairie root mass over six years (represented in days after establishment). Pink lines are the fit for each experimental plot and were used to make predictions.



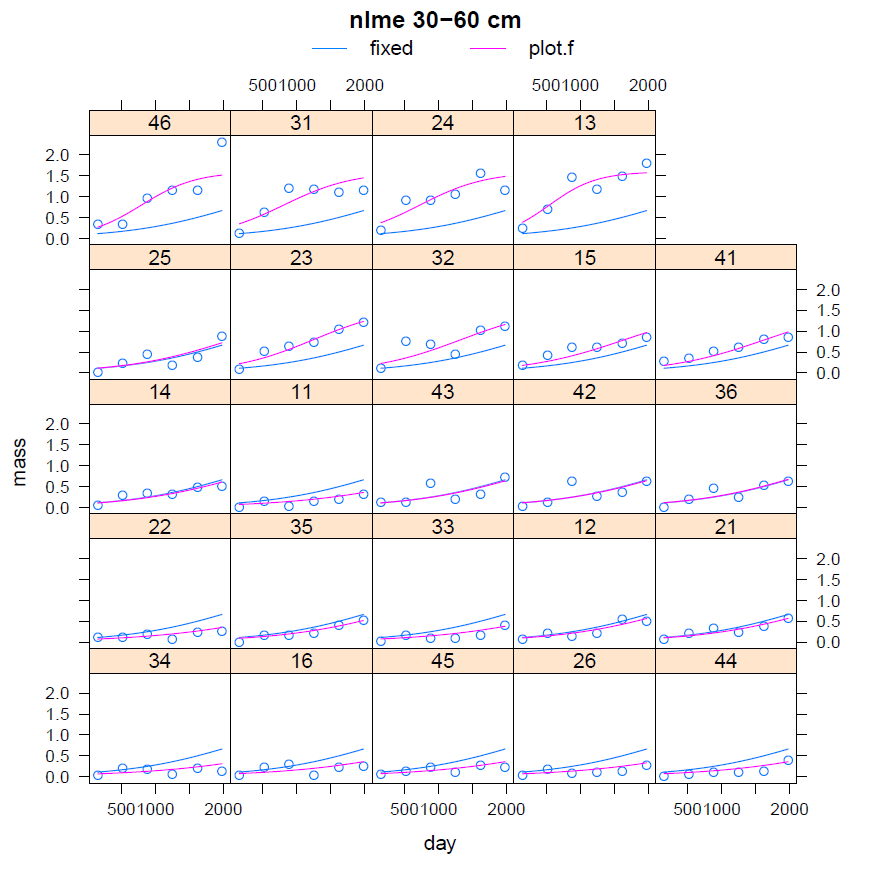
Appendix Fig 2. Fits of logistic curves to 0-5 cm row-crop root mass over six years (represented in days after establishment). Pink lines are the fit for each experimental plot and were used to make predictions.



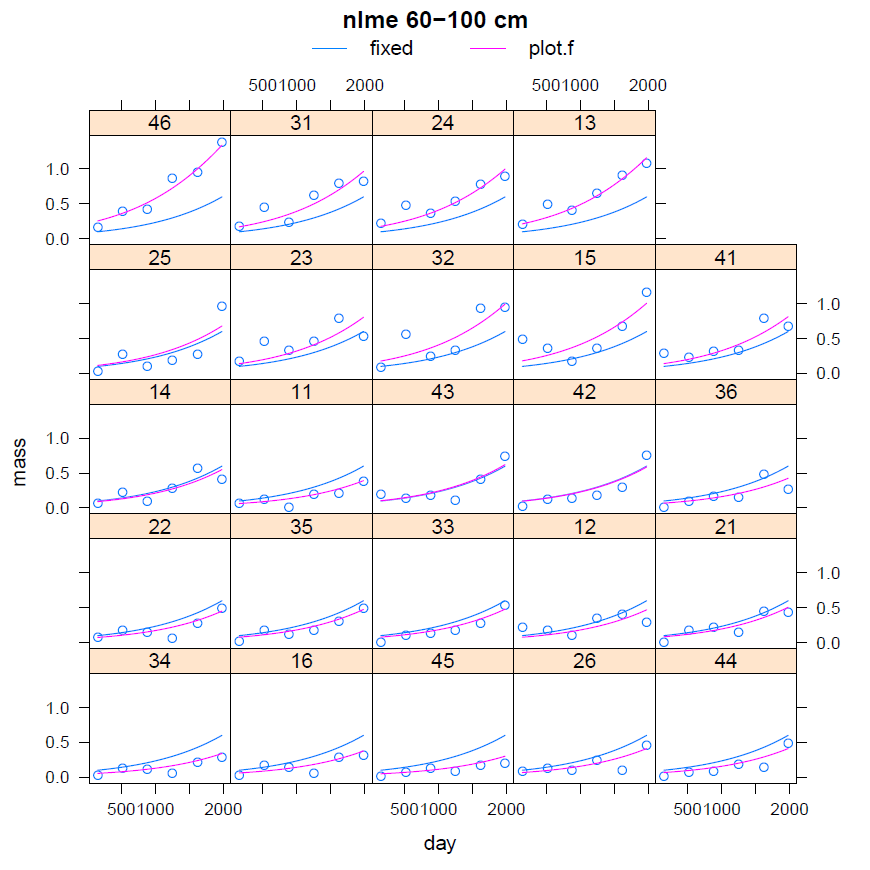
Appendix Fig 3. Fits of logistic curves to 5-15 cm root mass over six years (represented in days after establishment). Pink lines are the fit for each experimental plot and were used to make predictions.



Appendix Fig 4. Fits of logistic curves to 15-30 cm root mass over six years (represented in days after establishment). Pink lines are the fit for each experimental plot and were used to make predictions.



Appendix Fig 5. Fits of logistic curves to 30-60 cm root mass over six years (represented in days after establishment). Pink lines are the fit for each experimental plot and were used to make predictions.



Appendix Fig 6. Fits of logistic curves to 60-100 cm root mass over six years (represented in days after establishment). Pink lines are the fit for each experimental plot and were used to make predictions.